

Comparative biochemistry studies of the energy metabolism
in large animals

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2016

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Introduction

1. Clinical Biochemistry in Animals

For a long time, many different animals have shared their lives with humans. Since the animals live in a different environment from their original natural habitat, health monitoring is always important in terms of their safety and protection from diseases. It is especially important to diligently monitor for any abnormalities of their health status and any changes in their behavior in the early stages to obviate any health problems. Given this situation, understanding their normal health condition is one of the essential challenges for building a better living experience.

In this study, we focused on the lipid and energy metabolism in large animals. Since blood collection can be easily performed during routine health examinations, plasma metabolite values can be helpful to understand an animal's health condition. Like humans, many major diseases are related to metabolic conditions, so investigating metabolic biomarkers is beneficial for both animals and humans. Thus, monitoring and understanding the relevance of hematological data is crucial for health management of animals.

Lipoproteins play valuable role in energy and lipid metabolism of animals, and these patterns correlate with various metabolic changes. Since lipoprotein density profiles among animals are varied, it is possible to characterize different species by fraction patterns (Terpstra et al. 1982). As such, there are 2 main lipoprotein groups: HDL and LDL dominant mammals. The HDL dominant group consists of

dogs, cats, and horses; whereas the LDL dominant group includes humans, pigs, and rabbits (Chapman 1986). Cholesterol lipoprotein profiles can be useful to evaluate metabolic status such as overweight and obesity in dogs and cats (Mori et al. 2011; Muranaka et al. 2011). Thus, it is crucial to understand standard normal cholesterol lipoprotein patterns in order to detect any abnormalities in metabolic condition of animals.

Increased enzyme activity within the malate-aspartate shuttle appears to produce efficient energy metabolism in animal tissues (Arai et al., 1998), with malate-aspartate shuttle activity modulating according to variations in energy metabolism needs (Arai et al., 2001). Malate dehydrogenase (MDH) is necessary for gluconeogenesis and lipogenesis, and plays a significant role in the malate-aspartate shuttle (Setoyama et al., 1988). This transports cytosolic NADH into mitochondria, followed by oxidative ATP production (Hedekov et al., 1987). Alternately, lactate dehydrogenase (LDH) converts pyruvate to lactate, consuming cytosolic NADH. Glucose is thought to be metabolized to pyruvate during cytosolic glycolysis, and is the main energy source in most mammals (Lauris et al., 1966). Each animal species has specific values for plasma and leukocyte MDH, LDH enzyme activities, and MDH/LDH activity (M/L) ratio (Arai et al., 2003). Furthermore, Washizu et al. (2001) reported that cytosolic M/L ratio is considered to be a useful indicator to evaluate metabolic states in animal tissues.

2. Metabolic and Age-related Disorders in Animals

In recent years, progress of veterinary science has contributed to the longevity of companion animals, such as dogs and cats. However, along this development, the number of metabolic and age-related disorders in animals has been rapidly increasing. Metabolic disorders, such as diabetes and hyperlipidemia, have become one of the major health problems in companion animals. In some cases, the animal owners overlook the abnormal state until it is in a terminal condition. Additionally, heart, liver and kidney problems, as well as cancers, are representative examples of age-related disorders. Similarly to humans, these disorders can occasionally lead to comorbid conditions and become a more complicated status to treat. Therefore, diagnosis in the early stages and prevention of these disorders need urgent attention.

Specifically, obesity is a primary risk condition for metabolic and age-related disorders (Bray and Bellanger 2006). Interestingly, obesity-induced or obesity-associated diseases appear to arise from inflammation, and this type of inflammation contributes to the development of many chronic conditions, including diabetes, liver diseases, and some forms of cancer (Lukens et al., 2011; Stienstra et al., 2012; Gregor and Hotamisligil 2011; Osborn and Olefsky 2012). Recently, bidirectional immunological-metabolic crosstalk has made a great contribution and provided a new aspect to the relationship between health and disease in the human medical research area. Metabolic systems range from energy use to immunological defense. Metabolic dysfunction resulting from an obese condition is tightly connected to a variety of chronic diseases.

3. Significance of Comparative Study

Comparative studies are helpful because this knowledge can offer critical information, leading to establishing the basic standard data and discovering the distinct features of the animals. It is also advantageous to use this knowledge for other fields, including the human medical field.

According to the Japanese Association of Zoos and Aquariums (JAZA), we have placed a variety of animals in over 150 zoos and aquariums (registered in JAZA, 2015) for educational and/or breeding purposes throughout Japan. This captivity has benefited our research and understanding of their biology for a long time. However, diseases and conditions of the captive animals are relatively difficult to find or determine because they are often a rare species. Especially, research on rare species, such as aquatic mammals, is often problematic, thus information about the animals is scant, and the standard values of various blood biomarkers are not established in many cases. Since many different animals are bred in captivity, comparing this knowledge to common and familiar animals can be a valuable strategy in order to investigate and collaborate the information with other rare animal species.

Since the prevalence of overweight and obesity in animals has been increasing in recent years, learning about the metabolic systems in animals is imbued with significance. A better understanding of metabolism patterns can lead to more efficient management and disease prevention strategies for these animals.

Even though some animal species are closely related, each species has specific

features due to their environment or food habits. Comparing different species can accentuate the differences from other animals and provide useful clues to understand the animals. From this information, we can speculate or establish standards for the animals, and often apply the knowledge to human medical fields. For instance, it was demonstrated that metabolic responses in dolphins are consistent with human type 2 diabetes, thus dolphins can be used as animal models for this diseases (Venn-Watson et al. 2011). Dolphins are unique animals, sharing some similarities with humans and primates, such as glucose metabolism and brain size (Craik et al. 1998; Marino 1998), but there are differences as well. Therefore, it is of interest to research this animal in terms of the general metabolic system.

In this study, we investigated and compared metabolic systems in large animals. In captive condition, it is necessary to control and manage their feeding formula and physical activity, based on the individual health status of the animals. Since we previously researched various blood biomarkers of cows and horses, using the established data can increase the opportunities to give more pragmatic evidence of normal metabolism in other animal groups. We strongly believe that such comparative studies can be the very first step to a better health management of animals and humans.

Chapter 1

Cholesterol Lipoprotein Profiles of Large Animals

1-1 Introduction

Mammals have their own characteristics in glucose and lipid metabolism, reflecting in different plasma metabolite values (Sako et al. 2007). Understanding and monitoring the metabolic system in each animal species can lead to early detection and prevention of various metabolic-related diseases. In the veterinary field, obesity has become the popular health issue for companion animals (Laflamme 2012). Hyperlipidemia, commonly resulted from obesity, is the increased concentration of triglyceride (hypertriglyceridemia), cholesterol (hypercholesterolemia), or both in the blood (Watson and Barrie 1993; Ford 1996; Johnson 2005). Pathology of hyperlipidemia can be attributed to increased lipoprotein synthesis or mobilization or decreased lipoprotein clearance, and it can be primary (genetic or idiopathic) or secondary to other disease processes (Whitney 1992).

Lipoproteins have a valuable role in energy and lipid metabolism of animals, and these patterns correlate with various metabolic changes. Since lipoprotein density profiles among animals are varied, it is possible to characterize different species by fraction patterns (Terpstra et al. 1982). As such, there are 2 main lipoprotein groups: HDL and LDL dominant mammals. The HDL dominant group consists of dogs, cats, and horses; whereas the LDL dominant group includes humans, pigs, and rabbits (Chapman 1986). Cholesterol lipoprotein profile can be useful to

evaluate a metabolic status, such as overweight and obesity in dogs and cats (Mori et al. 2011; Muranaka et al. 2011). Thus, it is crucial to understand standard normal cholesterol lipoprotein patterns in order to detect any abnormalities in an animal's health condition.

Therefore, the aim of this study was to examine and compare plasma metabolite values and cholesterol lipoprotein profiles between dolphins, horses, and cows. As described above, each animal species has a different nutrition metabolic system, so it is intriguing to seek the similarity or the difference between these animal groups. Especially, since little is known about the health condition of marine mammals such as dolphins, a better understanding of their metabolism patterns can lead to the early prevention of various diseases. These 3 different animal species are similarly large animals, and have also been trained or domesticated by humans as well. Thus, horses and cows can serve as practical comparisons. Additionally, the basic information gained from comparative studies can provide profitable knowledge for establishing standards of animal basal metabolism.

1-2 Materials and Methods

Animals

Five captive bottlenose dolphins (*Tursiops truncatus*), 6 Thoroughbred riding horses, and 12 lactating Holstein cows were examined in this study. All animals were diagnosed to be healthy and exhibited no clinical signs for disease. The bottlenose dolphins were part of the population at Enoshima Aquarium in Kanagawa, Japan. The tank of the dolphin pool is outdoors with a total sea water

volume of 5000 m³ (45m x 25m, oval-shaped, with a depth of 3.5 to 5.5m) (Terasawa et al. 2005). The average monthly water temperature of the dolphin pool varied between 12.5±0.6°C in February to 27.3±1.4°C in August, and the average monthly air temperature ranged from 8.7±2.4°C in January to 27.9±2.2°C in August (Terasawa et al. 2005). The Thoroughbred riding horses were maintained and trained at Nippon Veterinary and Life Science University. The lactating Holstein cows were maintained at Koizumi Milk Farm in Tokyo, Japan.

Dolphins were fed 4 to 8 times per day, between 9:00 and 17:30, and consumed approximately 10 to 14 kg (mainly fish) daily. Horses were fed a daily diet of ~5 kg in total, 3 times per day, between 6:00 and 16:00. Cows were given 29 to 32 kg of feed daily, administered 4 times daily between 5:30 and 10:00. All animals involved in this study lived in captivity. Approval for this study has been given by the Nippon Veterinary and Life Science University Animal Research Committee.

Sample Collection

Blood sampling from dolphins was conducted on a monthly basis, as a part of their routine physical exams at Enoshima Aquarium. Briefly, blood was taken from the tail fluke of dolphins by venipuncture, using a sterile 21 gauge disposable butterfly needle. Because the dolphins were trained for blood sampling and/or other routine physical exams, they can voluntarily display their tail or body when samples need to be collected. All dolphin blood samples were obtained 16 to 17 hours after their last feeding. Blood sampling from horses and cows was obtained from the jugular or caudal vein, respectively, early in the morning (5:00-7:00 am). The animals

showed no excitement or fear, when blood sampling was conducted, since they were accustomed to blood sampling. Blood samples from all animals were centrifuged at 3500 rpm for 10 min at 4°C in order to obtain plasma samples, which were subsequently frozen at -80°C until further use.

Plasma Metabolite Assays

Plasma glucose, triglycerides (TG), and total cholesterol (T-Cho) were measured using an AU680 auto analyzer (Beckman Coulter, CA, USA) with the manufacturer's reagents. Non-esterified fatty acid (NEFA) concentration was measured using a Wako NEFA-C test commercial kit (Wako Pure Chemical Industries, Inc., Tokyo, Japan).

Cholesterol Lipoprotein Profiling

Plasma cholesterol lipoprotein patterns were detected by the biphasic agarose gel electrophoresis method utilizing commercial Quickgel Lipo gels (Code No. J715, Helena Laboratories, Saitama, Japan). In brief, 30 μ L sample volumes were loaded into the dipping well of an Epalyzer 2 Electrophoresis Processing Analyzer (Helena Laboratories), of which 5-6 μ L was loaded onto the gel, and run with a 14 min set migration time at 250 V and at 20°C. After migration, the gels underwent a 15 min reaction time, followed by a 12 min and 30 s decolorizing and fixing time, respectively. Cholesterol lipoprotein fractions were assessed and analyzed using Edbank III analysis software (Helena Laboratories).

Statistical Analysis

Results are expressed as means \pm SD. Paired groups were compared using the Mann-Whitney U-test for data with non-normal distribution, and statistical significance was set at $P < 0.05$. All tests were performed using Sigmaplot analysis software (Sigmaplot 11.0, Build 11.0.007; Systat Software Inc., San Jose, CA).

1-3 Results

Blood Biochemistry Values

All values for the measured plasma metabolites for dolphins, horses, and cows are presented in Table 1-1. Plasma glucose level in dolphins was similar to that in horses, but significantly higher than that in cows (Mann Whitney U-test, $p < 0.05$). Dolphins also had significantly higher levels of TG and NEFA than either horses (TG: $p = 0.019$, NEFA: $p = 0.002$) or cows (TG: $p = 0.001$, NEFA: $p = 0.004$). Dolphin and cow T-Cho levels were significantly higher as compared to horses ($p = 0.002$).

Table 1-1. Plasma Metabolite Comparison between Dolphins, Horses and Cows

	Dolphins (n = 5)	Horses (n = 6)	Cows (n = 12)
Glucose (mg/dL)	107.00 \pm 16.20	100.67 \pm 4.84	71.36 \pm 5.05*
TG (mg/dL)	22.80 \pm 8.70	14.33 \pm 5.28*	8.14 \pm 1.46*
T-Cho (mg/dL)	195.20 \pm 61.20	73.50 \pm 2.88*	217.07 \pm 40.57
NEFA (mEq/L)	0.32 \pm 0.12	0.04 \pm 0.02*	0.09 \pm 0.030*

Values are presented as mean \pm SD.

*Denotes significant difference as compared to dolphins (Mann-Whitney U-test, $p < 0.05$).

Cholesterol Lipoprotein Patterns

Cholesterol lipoprotein patterns for all animal groups are shown in Figure 1-1. All animal groups showed HDL dominant patterns. Although all animal groups displayed different patterns, dolphins and horses showed a similarity with clear fraction of LDL in both groups. There was no clear LDL fraction in cows.

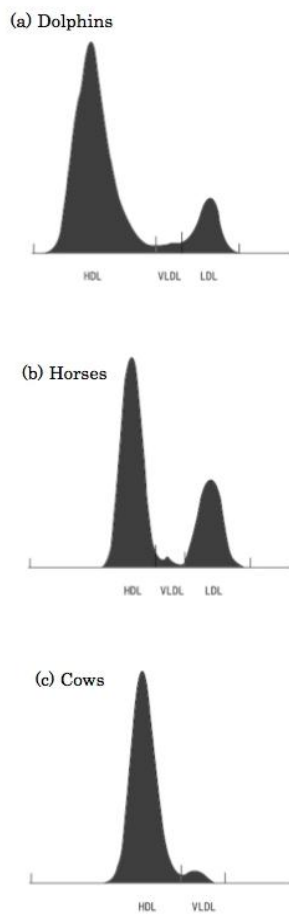


Figure 1-1. Comparison of cholesterol lipoprotein patterns between (a) dolphins, (b) horses, and (c) cows. Lipoproteins were separated by an electrophoretic technique and analyzed using Epalyzer 2 Electrophoresis Processing Analyzer (Helena Laboratories).

1-4 Discussion

As it is well known today, obesity is an increasingly encountered health problem, and it could increase the risks for serious diseases and health conditions. Prevalence of overweight and obesity is not only for human health problems, but also for animals (TvariJonaviciute et al. 2012; Martin et al. 2014; Robin et al. 2015). Since no reference ranges for plasma based parameters have been correlated with stages of obesity as of yet, objective and pragmatic biomarkers reflecting dysfunction of the metabolic state is needed in the veterinary field.

As it has been reported that obesity in dogs can result in aberration to cholesterol lipoprotein fraction profiles (Jericó et al. 2009), it is possible to understand the metabolic states of animals based on lipid and lipoprotein parameters. Moreover, it was suggested that analyzing serum lipid and lipoprotein values could characterize the different equine breeds and might be useful for evaluating metabolic states in horses (Asadi et al., 2006). It was also reported that the Shetland ponies had higher plasma TG and VLDL concentrations than their Thoroughbred counterparts (Watson et al., 1991). Therefore, understanding the metabolic system similarities and differences of the same animals group, can lead to more specific evaluation of each animal metabolic condition.

As the present data showed, cholesterol profile patterns between dolphins, horses and cows showed that all animal groups were classified into HDL dominant mammals, such as dogs and cats. Dolphins and horses also displayed clear LDL-Cho peak, which was not shown in cows, thus the cholesterol profile of dolphins is a closer resemblance to that of horses, rather than that of cows.

With dolphins, there is evidence showing that their health and environmental conditions might induce the changes in lipid metabolism. For instance, hyperlipidemia of captive Bottlenose dolphins during pregnancy (Terasawa and Kitamura, 2005), and seasonal changes in T-Chol value in captive Bottlenose dolphin sera (Terasawa et al., 2002) have been reported. Therefore, application of plasma lipid concentrations as diagnostic markers for metabolic disorders should be considered in relation to various factors inducing metabolic changes such as pregnancy, season and other considerable diseases. Moreover, daily diet is one of the most important factors that affect the plasma lipid concentrations and compositions. As the report in 2005 suggested that rice bran oil affects plasma lipid concentrations and lipoprotein composition in mares (Frank et al., 2005), specific food could affect the values in lipid metabolism. Further studies on the relationship between food compositions and lipid metabolism in dolphins are especially needed. Although there have been no reports, dolphins maintained at aquariums can sometimes become overweight. Determining the normal ranges of plasma metabolites and lipoprotein pattern, and what kind of aberrations in their metabolism can cause obesity, are crucial to manage their healthy metabolic state.

1-5 Conclusion

Evidence now indicates that understanding the characteristic in lipid and lipoprotein parameters of each animal group is necessary for maintaining healthy metabolism in animals. In addition, various factors can often alter the patterns, so investigating how they are involved in lipid metabolism can lead to a more specific

evaluation of an animal's metabolic state. Overall, monitoring of plasma lipid concentrations and the cholesterol profile in animals is useful to detect abnormal metabolic states in order to prevent various metabolic-related disorders.

Chapter 2

Comparison of Energy Metabolism and LDH Isoenzyme Patterns in Large Animals

2-1 Introduction

Monitoring and understanding the relevance of hematological data is crucial for health management of animals, especially those bred in captivity. Since little is known about the health conditions of marine mammals, it is very difficult to determine disease development, until it is in well advanced stages. A better understanding of metabolism patterns can lead to more efficient management and disease prevention strategies of these animals. Since blood collection can be easily performed, during routine health examinations, plasma metabolite values can serve as a useful index for early detection of diseases. With dolphins, it is rather difficult to detect health abnormalities, especially in the early stages, owing to the fact that standard normal plasma metabolite value ranges have yet to be established and little is understood thus far. Moreover, information about dolphin energy metabolism is scant so further developments will be of use to better understand basic dolphin basal metabolism.

Increased enzyme activity within the malate-aspartate shuttle appears to produce efficient energy metabolism in animal tissues (Arai et al. 1998), with malate-aspartate shuttle activity modulating according to variations in energy metabolism needs (Arai et al. 2001). Malate dehydrogenase (MDH) is necessary for gluconeogenesis and lipogenesis, and plays a significant role in the malate-

aspartate shuttle (Setoyama et al. 1988), transporting cytosolic NADH into mitochondria followed by oxidative ATP production (Hedeskov et al. 1987). Alternately, lactate dehydrogenase (LDH) converts pyruvate to lactate consuming cytosolic NADH. Glucose is thought to be metabolized to pyruvate, in cytosolic glycolysis, and is the main energy source in most mammals (Lauris and Cahill 1966). Each animal species has specific values for plasma and leukocyte MDH, LDH enzyme activities, and MDH/LDH activity (M/L) ratio (Arai et al. 2003). Furthermore, Washizu *et al.* (2001) reported that cytosolic M/L ratio is considered to be a useful indicator to evaluate metabolic states in animal tissues.

“Symmorphosis” as originally proposed by Taylor and Weibel (1981) states that biological design will be optimized, such that each structural element in a functional chain matches the maximal requirements of the overall functional system (Garland and Huey 1987). The concept was motivated by the authors’ beliefs that animals are built reasonably and their conviction that structural design is optimized because maintaining biological structures with their often high turnover rates is costly (Weibel and Taylor 1981). For an initial empirical test of symmorphosis, Weibel and Taylor focused on the mammalian respiratory system and proposed two comparative tests. First, for animals differing in size, the scaling of respiratory structures should parallel the scaling of maximal oxygen consumption. Second, for animals of similar size, differences in structural capacities should match differences in VO₂ max, the maximal rate of O₂ consumption attained during exercise of progressively increasing intensity. VO₂ max sets an upper limit to the intensity of work that can be sustained aerobically

for prolonged periods, and hence should be of considerable selective importance. As such, different species should reflect optimization and differences in oxidative energy metabolism, which would be reflected in muscle fiber (aerobic versus anaerobic) and metabolic profile differences between species.

Therefore, the aim of the present study was twofold. First, we sought to determine plasma metabolite profiles of healthy captive dolphins, and assess their energy metabolism state by examining plasma MDH and LDH activities, M/L ratio, and LDH isoenzyme patterns since such information is lacking in the literature. Second, we sought to compare the aforementioned parameters between healthy captive bottlenose dolphins and thoroughbred riding horses and lactating Holstein cows, in order to assess “Symmorphosis” regarding oxidative energy metabolism. Horses and cows may serve as practical comparisons for the following reasons. First, horses and cows are similarly large animals as dolphins, and have also been trained or domesticated by humans, as well. Second, because dolphins normally have relatively good motility, horses and cows can serve as good comparisons because horses are also quite active, whereas cows exhibit little physical activity; thereby highlighting differences in energy usage between all 3 species. Third, there is evidence demonstrating that dolphins may be closely related to even-toed ungulates, such as cows, pigs and camels (Shimamura et al. 1997). Lastly, our laboratory has previously reported on energy metabolism of horses and cows (Arai et al. 2003; Kimura et al. 2005; Li et al. 2012).

2-2 Materials and Methods

Animals

Five captive bottlenose dolphins (*Tursiops truncatus*), 6 Thoroughbred riding horses, and 12 lactating Holstein cows were included in this study. All animals were diagnosed to be healthy and exhibited no clinical signs for disease. The bottlenose dolphins were part of the population at Enoshima Aquarium in Kanagawa, Japan. The tank size, seawater volume, air and water temperatures of the dolphin pool at Enoshima Aquarium have been previously presented (Terasawa et al. 2005). Thoroughbred riding horses were maintained and trained at Nippon Veterinary and Life Science University. The lactating Holstein cows were maintained at Koizumi Milk Farm in Tokyo, Japan. Dolphins were fed 4 to 8 times per day, between 9:00 and 17:30, and consumed approximately 10 to 14 kg (mainly fish) daily. Horses were fed a daily diet of ~5kg in total, 3 times per day, between 6:00 and 16:00. Cows were given 29 to 32 kg of feed daily, administered 4 times daily between 5:30 and 10:00. All animals involved in this study lived in captivity.

Regarding physical activity, in general, dolphins at the Enoshima Aquarium were active as compared to other animal groups, constantly moving in and around the tank, due to training or shows in the daytime. Alternatively, horses at the university were usually immobile during the day, standing still, except for their daily exercise regiment, lasting 1 to 2 hours daily. Lastly, cows at the Koizumi Milk Farm were immobile, usually standing still. The cows were able to supply approximately 30 mL of milk per day. Approval for this work has been given by

the Nippon Veterinary and Life Science University Animal Research Committee.

Sample Collection

Blood sampling from dolphins was conducted on a monthly basis, as a part of their routine physical exams at Enoshima Aquarium. Briefly, blood was taken from the tail fluke of dolphins by venipuncture, using a sterile 21 gauge disposable butterfly needle. Because the dolphins were trained for blood sampling and/or other routine physical exams, they can voluntarily display their tail or body when samples need to be collected. All dolphin blood samples were obtained 16 to 17 hours after their last feeding. Blood sampling from horses and cows was obtained from the jugular or caudal vein, respectively, early in the morning (5:00-7:00 am). The animals showed no excitement or fear, when blood sampling was conducted, since they were accustomed to blood sampling. Blood samples from all animals were centrifuged at 3500 rpm for 10 min at 4°C in order to obtain plasma samples, which were subsequently frozen at -80°C until further use.

Plasma Metabolite Assays

Plasma biochemistry analysis on alanine aminotransferase (ALT), alkaline phosphatase (ALP), aspartate aminotransferase (AST), blood urea nitrogen (BUN), creatinine, glucose, lactate dehydrogenase (LDH), total cholesterol (T-Chol), total protein (TP), and triglycerides (TG) was performed using an AU680 auto analyzer (Beckman Coulter, CA, USA) with the manufacturer's reagents. Non-

esterified fatty acid (NEFA) concentration was measured using a Wako NEFA-C test commercial kit (Wako Pure Chemical Industries, Inc., Tokyo, Japan).

Enzyme Activity Assays and LDH Isoenzyme Profiling

The activities of malate dehydrogenase (MDH) and lactate dehydrogenase (LDH) in plasma were measured as previously described (LDH–Kaloustian et al. 1969 and MDH–Bergmeyer and Brent 1974). All enzymatic activities were measured between 24-26 °C and expressed as U per liter of plasma. The enzyme unit (U) was defined as 1 μ mol of substrate degraded per min. The M/L ratio was calculated as MDH activity divided by LDH activity. Plasma LDH isoenzyme patterns were detected by the biphasic agarose gel electrophoresis method utilizing commercial Quickgel LD gels (Helena Laboratories, Saitama, Japan). In brief, 30 μ L plasma sample volumes were loaded into the dipping well of an Epalyzer 2 Electrophoresis Processing Analyzer, of which 5-6 μ L was loaded onto the gel, and run with a 13 min set migration time at 240 V and at 15 °C. After migration, the gels underwent a 14 min reaction time, followed by a 12 min and 30 s decolorizing and fixing time, respectively. LDH fractions were assessed and analyzed using Edbank III analysis software (Helena Laboratories).

Statistical Analysis

Results are expressed as means \pm SD. Paired groups were compared using the Mann-Whitney U-test for data with non-normal distribution, and statistical significance was set at $P < 0.05$. All tests were performed using Sigmaplot analysis

software (Sigmaplot 11.0, Build 11.0.077; Systat Software Inc., San Jose, CA).

2-3 Results

Plasma Metabolite Assays

All values for the various measured plasma metabolites, for all animal groups, are shown in Table 2-1. There was no significant difference in glucose levels between dolphins and horses; however dolphin glucose level was significantly higher as compared to that of cows (Mann Whitney U-test, $p < 0.05$). Dolphins also displayed a significantly higher level of TG, than either horses ($p = 0.019$) or cows ($p = 0.001$). T- Cho levels in dolphins and cows were both significantly higher than that of horses ($p = 0.002$). NFFA in dolphins was also significantly higher than that of either cows ($p = 0.004$) or horses ($p = 0.002$). Lastly, dolphins had a significantly higher level of ALP as compared to either cows ($p = 0.001$) or horses ($p = 0.002$).

Table 2-1. Comparison of Blood Biochemistry Values of Dolphins, Horses, and Cows

	Dolphins (n = 5)	Horses (n = 6)	Cows (n = 12)
Glucose (mg/dL)	107.00±16.20	100.67±4.84	71.36±5.05*
TG (mg/dL)	22.80±8.70	14.33±5.28*	8.14±1.46*
T-Cho (mg/dL)	195.20±61.20	73.50±2.88*	217.07±40.57
BUN (mg/dL)	50.48±9.48	17.40±3.17*	9.18±2.02*
Creatinine (mg/dL)	1.22±0.23	1.36±0.13	0.60±0.09*
TP (g/dL)	7.80±0.56	6.23±0.340	7.73±0.68
LDH (IU/L)	438.40±34.06	216.67±58.84*	902.21±131.37*
AST (IU/L)	174.52±34.16	275.05±33.25*	66.60±8.40*
ALT (IU/L)	49.00±15.22	6.50±1.64*	30.50±4.80*
ALP (IU/L)	1344.80±661.55	255.50±41.29*	146.64±78.94*
NFFA (mEq/L)	0.32±0.12	0.04±0.02*	0.09±0.03*

Values are presented as mean±SD.

*Denotes significant difference as compared to dolphins (Mann-Whitney U-test, p<0.05).

MDH, LDH, and M/L Ratio

A comparison of plasma MDH and LDH activities, in addition to M/L ratio between all 3 animal groups is displayed in Table 2-2. Although no significant difference in MDH activity was noted between any of the groups, LDH activity in dolphins was significantly (Mann Whitney U-test, p<0.05) higher than that of horses (~2 fold), and lower than that of cows (~65% lower), respectively. As a result, M/L ratio in each animal species was different from one another (Mann Whitney U-test, p<0.05). Horses demonstrated the highest M/L ratio (0.79), followed by dolphins (0.67) and cows (0.15), respectively.

Table 2-2. Comparison of MDH and LDH Activities and M/L Ratio in Plasma of Dolphins, Horses, and Cows

	Dolphins (n = 5)	Horses (n = 6)	Cows (n = 12)
MDH	134.51±90.17	69.24±17.68	90.00±20.69
LDH	199.00±27.63	89.05±17.66*	604.21±184.58*
M/L ratio	0.67±0.13	0.79±0.15	0.15±0.03*

Values are presented as mean±SD.

*Denotes significant difference as compared to dolphins (Mann-Whitney U-test, $p < 0.05$).

LDH Isoenzyme Patterns

LDH isoenzyme patterns for all animal groups are presented in Figure 2-1.

Although all animal groups displayed significant differences in isoenzyme pattern distribution, dolphins and horses demonstrated a similarity in LDH isoenzyme pattern with LDH-3 predominating, and LDH-2 and -3 making up 50% of total plasma LDH, in both species (Table 2-3). The isoenzyme pattern in dolphins, in descending % amount of total LDH, was LDH-3 (39.62±1.07%), LDH-2 (29.56±1.43%), LDH-4 (17.18±1.78%), and LDH-1 (10.46±0.97%). Alternately, the isoenzyme pattern in horses, in descending amount, was LDH-3 (31.55±2.20%), LDH-1 (29.88±3.02%), LDH-2 (25.30± 2.63%), LDH-4 (10.67±0.99%), and LDH-5 (2.60±0.95%). Unlike both dolphins and horse, LDH-1 (47.07±1.56%) was the most dominant isoform in cows, followed by LDH-2 (27.77±0.78), LDH-3 (15.45±1.11), LDH-4 (5.60±0.50), and LDH-5 (4.12±0.78).

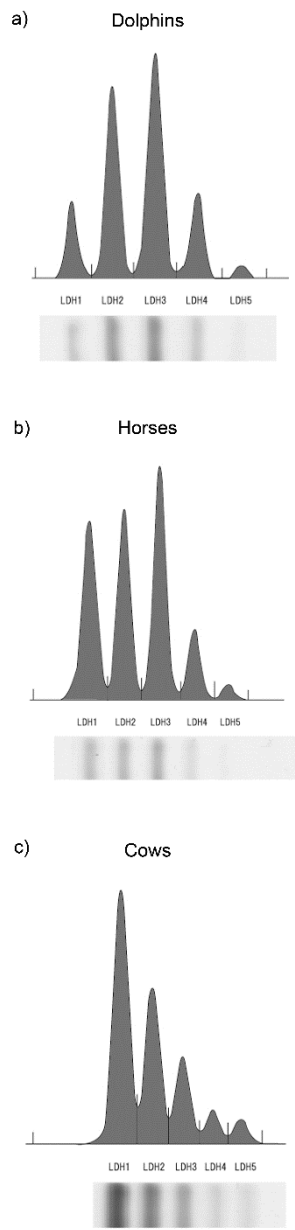


Figure 2-1. Comparison of plasma LDH isoenzyme patterns between cows, horses and dolphins. LDH isoenzymes were separated by an electrophoretic technique and analyzed using Epalyzer 2 Electrophoresis Processing Analyzer (Helena Laboratories).

Table 2-3. Comparison of LDH Isoenzyme Distribution between Dolphins, Horses, and Cows

	Dolphins (n = 5)	Horses (n = 6)	Cows (n = 12)
LDH1 (%)	10.46±0.97	29.88±3.02	47.07±1.56
LDH2 (%)	29.56±1.43	25.30±2.63	27.77±0.78
LDH3 (%)	39.62±1.07	31.55±2.20	15.45±1.11
LDH4 (%)	17.18±1.78	10.67±0.99	5.60±0.50
LDH5 (%)	3.18±0.63	2.60±0.95	4.12±0.78

Values are presented as mean±SD.

2-4 Discussion

“Symmorphosis” as originally proposed by Taylor and Weibel (1981) dictates that there should be a symbiosis between form and function for every animal species. In general, dolphins are constantly moving, hence their muscle fiber and metabolic profiles should reflect that species, as opposed to cows (do not run around at all) and horses (in between). Therefore, a difference in aerobic (endurance) vs anaerobic muscle fiber profiles will have a great impact on ATP usage, in addition to general activity levels for example. Two fundamental reactions resynthesize ATP (Duren 1998): 1) Oxidative phosphorylation, breaking down carbohydrates, fats and protein, in the presence of oxygen, producing energy (ATP). The involvement of oxygen qualifies this as an aerobic reaction. 2) Glycolysis, breaking down glucose or glycogen into lactic acid. This reaction does not use oxygen and is considered anaerobic. There are several factors which will determine both the choice of fuel and the pathway used to generate ATP. These factors include: muscle fiber type, the speed and duration of exercise, type of feed provided, and animal

fitness.

M/L ratio appears to be one potential useful indicator of oxidative metabolism in animals since it can indirectly measure ATP synthesis/regeneration rate. A ratio of 1 would be ideal, since MDH enzyme activity to generate ATP is equally balanced by LDH enzyme activity to regenerate NADH into NAD⁺ for additional energy production. Therefore, ATP is resynthesized at the same rate at which it is being used. However, since it is possible to obtain similar ratios, with differing MDH and LDH activity levels, it is imperative that MDH and LDH levels are also analyzed, in context, in order to reflect whether elevated energy metabolism, including elevated ATP production, may be occurring in some tissues, such as muscle and liver. Plasma M/L ratio in horses (0.79) was greater than that of dolphins (0.67) or cows (0.15); however MDH and LDH enzyme activity levels in dolphins was ~1.9 and ~2.2x higher, respectively than that of horses; thereby indicating greater overall energy usage in dolphin than horse tissues. LDH, aldolase and phosphoglucomutase levels are higher in dolphin than in other mammalian skeletal muscles; pyruvate kinase is the only glycolytic enzyme in lower concentration in dolphin muscle (Storey and Hochachka 1974). Therefore, although horses appear to use energy more efficiently than dolphins, there is greater energy usage or metabolism by dolphins as compared to horses, as evidenced in higher overall plasma MDH and LDH activity levels.

The horse has three basic types of muscle fiber: Type I, IIA and IIB, which differ in contractile and metabolic characteristics (Duren 1998). Type I fibers are slow-contracting fibers, while Types IIA and IIB are fast-contracting. In addition, Type

I and IIA fibers have a high oxidative capacity and thus can utilize fuels aerobically while Type IIB fibers have a low aerobic capacity and depend on anaerobic glycolysis for energy generation. All three fiber types store glycogen, while only Types I and IIA have significant triglyceride storage. The speed of muscle contraction determines how fast the animal is able to move, and the faster an animal moves the greater the ATP requirement (Duren 1998). While walking, muscle contraction occurs very slowly and relatively small amounts of ATP are expended. During this type of exercise, Type I fibers are primarily recruited and energy generation is entirely aerobic, with muscle burning predominantly fat. As speed increases from a walk to a trot to a canter, Type I fibers alone are no longer capable of contracting rapidly enough to propel the horse. At this point, Type IIA fibers are also recruited. These fibers are also aerobic, but they use a combination of glycogen and fat for energy generation. Glycogen (glucose) can be metabolized twice as fast as fat for ATP generation, and as speed increases, fat becomes simply too slow a fuel for energy generation. As the horse increases speed to a fast gallop, Type IIB fibers are recruited and energy generation no longer remains purely aerobic. At these speeds, the requirement for ATP has exceeded the ability of the horse to deliver enough oxygen to the muscle to produce the energy by aerobic means. Anaerobic glycolysis takes over as a rapid metabolic pathway to generate ATP. Anaerobic glycolysis results, however, in lactic acid accumulation, and fatigue soon develops as the pH in the muscle begins to fall.

Therefore, in the case of the riding horses used in our study, the majority of energy usage appears to be aerobic in origin due to the type of physical activity and muscle

profile involved. These facts would support the greater plasma M/L ratio in horses (0.79) as opposed to dolphins (0.67) since M/L ratio may be measuring oxidative ATP usage/regeneration by aerobic means only. Because dolphins are constantly moving, their energy usage may exceed that which can be produced by aerobic means only, and therefore M/L ratio does not take ATP generation by anaerobic means into account.

To further support the idea of increased energy usage or metabolism in dolphins as opposed to horses, dolphins demonstrated significantly higher levels of plasma lipid metabolites, such as triglycerides (TG, 0.5x greater), total cholesterol (T-Cho, 2.5x greater) and Non-esterified fatty acid (NEFA, 8x greater) as compared to horses. This may be indicative of dolphins having more capacity for lipolysis and lipid utilization as an additional energy source for aerobic ATP usage and regeneration, since they require more energy, and may have higher adipose stores as compared to horses (Rommel and Lowenstine 2001; Kasamatsu et al. 2009). The difference in adipose stores between horses and dolphins may be possibly related to the type and quality of their diets (Asper et al. 1990; Bossart et al. 2001; Kasamatsu et al. 2001). In addition, seasonal variation of plasma T-Cho and free fatty acid levels in dolphins has been reported with T-Cho and free fatty acid levels being inversely correlated with blubber thickness (Terasawa et al. 2002; Williams et al. 1992), with free fatty acid most likely being released from and deposited to the blubber in warm seasons and cooler winter months, respectively (Williams et al. 1992).

Additionally, dolphins exhibited the highest amount of plasma ALP amongst the animal species, being ~5x and ~9x greater than in horses and cows respectively. High levels of plasma ALP in dolphins have been previously observed in young and growing dolphins, with a significant decrease in concentration with increasing age (Fair et al. 2006).

Cows, on the other hand, demonstrated the lowest level of oxidative metabolism efficiency by aerobic means amongst all animal groups. Thus their aerobic energy usage may not be as efficient or may significantly differ from dolphins and horses, which would be expected according to “Symmorphosis” (Taylor and Weibel 1981). Since dolphins and horses are considered to be more active than cows, the increase in energy usage efficiency by aerobic means, of these 2 animal species could possibly be attributed to increased motility as compared to cows. In addition, since the cows are under a lactating state, energy usage by the animal is very different as opposed to a non-lactating state. Lactation in many species, particularly dairy cattle, which have been specifically bred for high milk production, requires a dramatic shift in metabolism redirecting a significant portion of ingested energy and metabolites to milk synthesis (Bell and Bauman 1997). In cattle, hepatic gluconeogenesis provides the majority of glucose required for mammary lactose synthesis. The liver also breaks down non-esterified fatty acids (NEFA) as an additional energy source if there is insufficient acetate and volatile fatty acids (VFA) coming from digestion. In addition, glucose metabolism of muscle and adipose tissue is also modulated during lactation, with gluconeogenesis being down-regulated in muscle tissue, and a shift in the ratio of lipogenesis to lipolysis

in adipose tissue, by hormones such as bovine somatotropin that stimulates galactopoiesis. In pregnant animals, these responses are exaggerated by moderate undernutrition and are mediated by reduced tissue sensitivity and responsiveness to insulin, associated with decreased tissue expression of the insulin-responsive facilitative glucose transporter, GLUT4. Peripheral tissue responses to insulin remain severely attenuated during early lactation but recover as the animal progresses through mid lactation. Therefore, the low level of energy metabolism, reflected by plasma M/L ratio, observed in lactating cows is indicative of the high energy requirement redirected for lactation instead of to the animal.

Interestingly though, cows demonstrated the highest level of plasma LDH activity amongst all animal species, being ~3x and ~6.8x greater than dolphins and horses, respectively. Because lactating Holstein cows require more energy for lactation, LDH enzyme activity may be increased, in many tissues, as compared to non-lactating cows. In addition, rumen bacterial fermentation, due to lactate-producing organism such as *Lactobacillus acidophilus* could also affect LDH value in cows (Boyd et al. 2011).

Different animal species exhibit differing characteristic plasma LDH isoenzyme patterns. In addition, each tissue has a characteristic composition of isoenzymes (Dawson et al. 1964). LDH is a tetrameric molecule made of four subunits of the parent molecules, and has five isoenzymes: LDH-1 (H₄), LDH-2 (H₃M₁), LDH-3 (H₂M₂), LDH-4 (H₁M₃) and LDH-5 (M₄) (Dawson et al. 1964). The present forms are termed H (heart) and M (muscle), for the organs from which they are obtained. Tissues containing a preponderance of LDH-1, with a lower K_m value for pyruvate,

usually function under aerobic conditions; whereas those containing a preponderance of LDH-5, function under relatively anaerobic conditions (Latner et al. 1966).

Although all animal groups displayed significant differences in isoenzyme pattern distribution, dolphins and horses demonstrated a similarity in plasma LDH isoenzyme pattern with LDH-3 predominating. However, the predominance patterns also reflect “Symmorphosis” of each species in that LDH-3,-2, and -4 predominated, in decreasing amount, in dolphins, which may often experience both anerobic and anaerobic energy utilization simultaneously, while LDH-1 and LDH-3 were equally predominant, with LDH-2 following in horses, which can also experience both aerobic and anaerobic energy utilization, however not simultaneously usually, and lastly LDH-1, -2, and -3 were predominant in cows, which will tend to only experience aerobic energy utilization, due to their lack of general mobility. As such, H type isoenzymes were relatively predominant in all of the animal species plasma examined in this study.

This study had several limitations. First, a small number of healthy dolphins and horses were available from the Enoshima Aquarium and the University Horse Riding club, respectively; therefore we had a limited number of animals in our study. Moreover, only one blood sample was drawn, per dolphin or horse, resulting in low statistical power, hence results and conclusions should be interpreted with care. Second, because the animals involved in the study were randomly selected, there was no age or gender matching performed. Aging can have an impact on energy consumption, however it is still uncertain whether it is influential enough

to affect enzyme activities. For example, higher MDH activities, in skeletal muscle of older men, undergoing endurance training, have been reported (Suominen and Heikkinen 1975). Nonetheless, investigating for alternation in enzyme activities may be more appropriate if measured against age and gender matched animal groups. Three, postprandial influence might have had an impact on plasma metabolite values such as glucose and triglycerides. All blood samples, except from dolphins, were collected within 5 h postprandially, hence postprandial influence cannot be eliminated. In general, fasting is recommended before lipid screening because of the theoretical dynamic changes that can occur in test results for some lipid components, however this study focused on energy metabolism, therefore postprandial influence would be limited. Fourth, all animals involved in this study lived in captivity, and it is critical for this study to examine the animals in captivity. Particularly, blood samples collected from wild dolphins may affect the data by several reasons such as their handling stress and their diet (Terasawa et al. 1999). Fifth, there was no standardized accounting or estimate of the activity and energy expenditure per day for the different animals used in our study. As such, no direct comparison could be made in terms of actual energy expenditure between the 3 different animal species.

2-5 Conclusion

In conclusion, dolphins appear to have the greatest level of oxidative energy metabolism amongst horses and cows, due to having the greatest levels of plasma MDH activity. In addition, dolphin energy production/usage efficiency was second (M/L ratio = 0.67) behind that of horses (M/L ratio = 0.79). Overall, these results suggest that dolphins may possibly generate more energy than horses, especially for increased mobility in the water; however all of the produced energy is not utilized for some reasons, such as smaller tank size or other environmental limitations. However, cows demonstrated the highest plasma LDH activity amongst all animal species. This may have been attributed to their lactating state. Although all animal groups displayed a different plasma LDH isoenzyme pattern distribution, dolphins and horses demonstrated a similarity with LDH-3 isoenzyme predominating in plasma, as opposed to LDH-1 in cows, which would reflect “Symmorphosis” of these 2 species and their aerobic/anaerobic energy metabolism needs.

Overall, plasma MDH and LDH activity levels, M/L ratio, and plasma LDH isoenzyme pattern can all be useful indicators for better understanding oxidative energy metabolism and monitoring of captive animals' health. As it is not easy to obtain tissue samples from animals, the development of blood indicators for evaluating whole body metabolic state is necessary, and further research is required and should be pursued.

Chapter 3

The Aging Effect on the Metabolic System in Riding Horses

3-1 Introduction

In recent years, progress of veterinary medicine has contributed to the longevity of companion animals. However, along this development, age-related disorders in animals have become a popular topic for clinical practice. As it is well known today, since a metabolic system is subject to change with aging, the number of metabolic disorders in animals has been rapidly increasing as well. In particular, prevalence of overweight and obesity has increased in dogs, cats, and horses (Tvariionaviciute et al. 2012; Martin et al. 2014; Robin et al. 2015). In general, obesity and overweight are caused by excess calorie intake and physical inactivity. It should be noted that the influence of physical activity is a critical factor for anti-obesity and/or anti-aging. Although domestic dogs and cats tend to decrease physical activity with aging, often times leading to an overweight or obese condition, the health of most riding horses is maintained with sufficient quality of motility.

Adiponectin (ADN), the most abundant protein secreted from adipose tissue, is commonly known for its involvement in obesity-related disorders, such as diabetes and insulin resistance. ADN regulates glucose metabolism and insulin sensitivity through AMP-activated protein kinase (AMPK) (Yamauchi et al. 2002). AMPK plays a crucial role in the regulation of energy metabolism, and it has been demonstrated that exercise acutely increases AMPK activity, as the beneficial effects of exercise were suggested (Ruderman et al. 2013). Since the horses in this

study had daily exercises, it is intriguing to investigate their plasma ADN and AMPK levels, and the connection with their energy metabolism. As shown in our previous reports, MDH/LDH activity ratio (M/L ratio) is considered to be a useful indicator to evaluate metabolic states in animal tissues (Hirakawa et al. 2012). We utilized this method to evaluate energy metabolism in horses.

Furthermore, it has been widely accepted that oxidative stress generated by free radicals is associated with the aging process. Harman proposed “The free radical theory of aging”, which states that aging is the cumulative result of oxidative damage caused by free radicals (Harman 1956). Malondialdehyde (MDA) is frequently used as a biomarker for oxidative stress, and it is known that superoxide dismutase (SOD) activity reflects the response against oxidative stress. The purpose of this study is to investigate the various metabolic biomarkers between three different age groups in riding horses in order to analyze the aging effect on their metabolic system. Importantly, horses are very active animals, so we discuss whether their daily physical activity can improve metabolic status, leading to a slowing aging process. Since blood collection can be easily performed during routine health examination, blood analysis can serve as a useful index for early detection and protection from metabolic-related and/or age-related diseases.

3-2 Materials and Methods

Animals

Nineteen riding horses maintained at Japan Horseback Riding Club (Saitama, Japan) were examined in this study (Figure 3-1). All horses were diagnosed to be

healthy and exhibited no clinical signs for disease. They were divided into three groups: young group (7-8 years old, average 7.6 ± 0.2 , $n=9$), middle-aged group (11-14 years old, average 12.4 ± 0.5 , $n=5$), and aged group (15-18 years old, average 15.8 ± 0.6 , $n=5$). The horses were fed 5.2 to 6.4 kg of hay cube, 3.0 to 4.0 kg of Italian ryegrass, 0 to 1.3 kg of wheat bran, and 0 to 1.8 kg of barley at 6:00 and 16:00 daily. The body weight of all the horses ranges from 450kg to 500kg. Their physical constitutions were assessed by the six-scale equine body condition score (EBCS), originally established by Carroll and Huntington scoring system (Carroll and Huntington 1988) with some modifications by Robin (Robin et al. 2015) as follows: very thin [1], thin [2], fair [3], good [4], fat [5], and very fat [6]. According to this scoring system, all the horses involved in this study were determined as EBCS 4 (good). Regarding physical activity, horses at Japan Horseback Riding Club had daily exercises 1 to 3 times per day, including walking at 100 to 110 m/min for 10 to 30 min, trotting at 200 to 220 m/min for 10 to 30 min, and cantering at 300 to 350 m/min for 15 min. The exercise amount of each horse depends on a riding lesson for an individual level such as beginner, intermediate, and senior courses. Ethical approval in this study was obtained from Japan Horseback Riding Club.



Figure 3-1. Representative horses in this study. Left: young horse (female, 7 years old, EBCS4). Right: aged horse (gelding, 18 years old, EBCS4).

Sample collection

Blood was withdrawn from the jugular vein of the horses into heparinized tubes. The horses showed no excitement or fear, when blood sampling was conducted. Plasma was recovered by centrifugation at 1200 g, for 5 min at 4°C, and stored at -80°C until further use.

Blood analysis

Plasma biochemistry analysis on glucose (GLU), total cholesterol (T-Cho), triglyceride (TG), total protein (TP), blood urea nitrogen (BUN), creatinine (CRE), alanine aminotransferase (ALT), aspartate aminotransferase (AST), and alkaline phosphatase (ALP) was performed using an auto analyzer (JCA-BM2250, JEOL Ltd., Tokyo, Japan) with the manufacture's reagents at Monolis Inc. (Tokyo, Japan).

The activities of malate dehydrogenase (MDH) and lactate dehydrogenase (LDH) in plasma were measured by previously reported methods (LDH–Kaloustian et al. 1969 and MDH–Bergmeyer and Brent 1974). The M/L ratio was calculated as MDH activity divided by LDH activity. Plasma non-esterified fatty acid (NEFA) concentration, malondialdehyde (MDA) concentration, and superoxide dismutase (SOD) activity were measured using a commercial kit: Wako NEFA-C test (Wako Pure Chemical Industries, Inc., Tokyo, Japan), NWLSSTM Malondialdehyde assay (Northwest Life Science Specialties, LLC, Vancouver, Canada), and NWLSSTM Superoxide Dismutase Activity Assay (Northwest Life Science Specialties, LLC, Vancouver, Canada), respectively. Plasma Insulin (INS) concentration and adiponectin (ADN) concentration were determined with a commercial ELISA kit: Lbis Rat T insulin kit (SHIBAYAGI Co., Gunma, Japan) and mouse/rat adiponectin ELISA kit (Otsuka Pharmaceutical Co., Ltd., Tokyo, Japan), respectively. Leukocytic AMP-activated protein kinase (AMPK) activity was measured using the CycLex AMPK Kinase Assay Kit (Medical & Biological Laboratories Co., Ltd., Nagano, Japan). Leucocytes were collected from 5 ml of buffy coat of whole blood. Cytosolic fractions of leukocytes were prepared and isolated according to the previously described method (Washizu et al. 1998) with some modifications.

Statistical analysis

Results are expressed as means \pm SE. Statistical analysis between the three groups was performed using the Mann-Whitney U test, and statistical significance was set at $P < 0.05$.

3-3 Results

All values for the various measured plasma biomarker levels for all the age groups are shown in Table 3-1. There was no significant difference in plasma metabolites between any of the groups. Plasma ADN concentration in the middle-aged group was significantly lower than that in the young group ($p=0.0136$), but the aged group displayed almost the same level of ADN as the young group. Leukocyte AMPK activities changed parallel to plasma ADN concentrations as both levels decreased.

Table 3-1. Comparison of the biochemical and energy metabolism parameters between young, middle-aged and aged groups of riding horse

	Young (n=9)	Middle-aged (n=5)	Aged (n=5)
Age (years old)	7.6 ± 0.2	12.4 ± 0.5	15.8 ± 0.6
GLU (mg/dL)	98.0 ± 2.6	100.2 ± 1.5	99.6 ± 2.4
TG (mg/dL)	16.3 ± 3.8	15.8 ± 2.2	13.6 ± 1.8
T-Cho (mg/dL)	70.4 ± 3.2	71.8 ± 4.9	77.6 ± 6.7
NEFA (µmol/L)	10.6 ± 4.0	45.2 ± 29.5	122.6 ± 86.3
MDA (µmol/L)	1.36 ± 0.05	1.45 ± 0.07	1.72 ± 0.17
SOD (U/mL)	4.2 ± 3.2	13.8 ± 7.0	15.3 ± 10.3
TP (g/dL)	6.5 ± 0.1	6.8 ± 0.2	6.6 ± 0.1
BUN (mg/dL)	18.0 ± 0.6	17.6 ± 0.8	20.0 ± 0.7
CRE (mg/dL)	1.1 ± 0.1	1.1 ± 0.1	1.2 ± 0.1
INS (ng/mL)	0.37 ± 0.08	0.51 ± 0.15	0.63 ± 0.27
ADN (µg/mL)	9.7 ± 1.4	4.8 ± 0.9*	8.2 ± 2.4
MDH (IU/L)	368.5 ± 45.6	385.6 ± 34.7	363.0 ± 17.2
LDH (IU/L)	202.9 ± 27.6	218.7 ± 12.0	182.5 ± 14.3
M/L ratio	1.89 ± 0.14	1.76 ± 0.11	2.02 ± 0.11
AST (IU/L)	265.1 ± 37.5	238.2 ± 14.3	227.2 ± 17.6
ALT (IU/L)	7.0 ± 1.2	6.4 ± 0.7	6.0 ± 0.5
ALP (IU/L)	254.3 ± 17.5	255.6 ± 24.5	242.0 ± 14.3
AMPK (ng/min mg/protein)	74.0 ± 15.6	40.6 ± 4.8	48.4 ± 11.2

Values are presented as mean ± SE.

*Denotes significant difference as compared to young group (Mann-Whitney U test, p<0.05).

3-4 Discussion

Consistent with our previous report (Kawasumi et al. 2015), plasma ADN level in the middle-aged group greatly decreased as compared to the young group (p=0.0136). However, in the aged group, this level appeared to increase again. ADN

is the most abundant protein secreted by white adipose tissue, and is one of the most thoroughly studied adipocytokines. As the protective action of ADN in obesity-linked diseases has been reported in numerous published articles, ADN has various beneficial effects on organs and tissues (Brochu-Gaudreau et al. 2010). It was also demonstrated that low plasma ADN levels are associated with obesity, metabolic syndrome, diabetes, and many other metabolic diseases in humans (Lu et al. 2008). Hence, it is advantageous to enhance ADN ability in order to prevent metabolic-related disorders. For instance, exercise is one of the important stimuli, and it was reported that ADN levels in older people could be improved by exercise (Lira et al. 2011). Interestingly, they also suggested that the improvement of metabolic profile in older people was time-dependent, indicating that proper training strategy is needed to achieve maximal beneficial effects. Therefore, in this study, the recovery of ADN level in aged horses was thought to be due to their continuous daily exercises, and possibly, applying proper exercise protocol modification to the middle-aged horses can even lead to keeping higher ADN levels throughout their lives. Moreover, we focused on the signaling pathways that mediate the metabolic effects of ADN. Since ADN regulates glucose metabolism and insulin sensitivity through AMPK (Yamauchi et al. 2002), AMPK was also measured in the present study. Recent studies revealed that AMPK and its related cellular signaling pathways play a critical role in exercise-mediated adaptations in the muscle (Sasaki et al. 2014), and AMPK activation in response to a number of stimuli including exercise is a potential target for metabolic diseases (Zhang et al. 2009). Although all the horses we examined in this study had daily exercises and

maintained good physical health, the result showed that AMPK levels in both the middle-aged and the aged groups tended to decrease as compared to the young group. Unlike ADN, there was no distinct recovery of AMPK in aged horses. In general, exercise can trigger AMPK activation in a time and intensity dependent manner (Jorgensen and Rose 2008), but a number of other stimuli are involved with AMPK activation as well. Therefore, other factors may affect AMPK levels.

Furthermore, M/L ratio, one of the useful indicators of energy metabolism, showed that there was no apparent decline between any of the groups. Because basal metabolism usually declines with aging, this result may imply that continuous daily exercises can ameliorate the aging process and can prevent age-related decline in energy metabolism. Additionally, exercise intensity and duration are also important enhancing factors for improvement of energy metabolism. It was demonstrated that longer and low-intensity exercise improves insulin action and plasma lipids more than shorter and high-intensity exercise (Duvivier et al. 2013). In contrast to race horses, all the riding horses in this study had moderate exercises on a daily basis, so their exercise protocol was advantageous for sustaining better energy metabolism. Although further studies should be required, continuous moderate exercise can be helpful for anti-obesity and/or anti-aging for horses.

Plasma NEFA levels tended to increase as the horses grow older. NEFA can induce inflammation through various mechanisms, and this chronic inflammation response is associated with dysfunction of metabolic and immune systems (de Heredia et al. 2012). However, NEFA levels in horses can be affected by several

factors, including duration and intensity of exercise (Li et al. 2012). Moreover, in this study, all the horses maintained good physical condition (EBCS 4), and there was no significant elevation in TG or T-Chol among all the age groups, so increased NEFA levels in middle-aged and aged horses may not be the result of lipid metabolism dysfunction. For further research, other triggers possibly affecting NEFA should be reviewed, and it is crucial to detect inflammatory changes induced by NEFA.

It has been commonly known that oxidative stress is associated with aging since Denham Harman proposed the Free Radical Theory of Aging in 1956 (Harman 1956). This theory states that aging is the cumulative result of oxidative damage caused by free radicals. Lipid peroxidation provoked by free radicals is possibly involved with the aging process because lipids are a major component of living organisms and can be the first target of free radicals (Praticò 2002). MDA is one of the final products of polyunsaturated fatty acids peroxidation in the cells, and overproduction of MDA indicates an increase in free radicals (Gawel et al. 2004). Therefore, MDA can be used as a marker of oxidative stress. The present study showed that there was no remarkable change in MDA concentrations between any of the groups. Moreover, SOD levels tended to increase in both the middle-aged and the aged groups as opposed to the young group. It might be suggested that SOD, which is an antioxidant enzyme, functioned and prevented from excess free radical formation. Interestingly, it was reported that low-intensity exercise can boost SOD activity and decrease MDA, but high-intensity exercise adversely affects the free radical scavenging ability of SOD, resulting in slowing the

reduction of MDA in rats (Li et al. 2015). Generally, exercise is important for health, but exercise intensity could be a key to improve the metabolic system more effectively. Therefore, the horses examined in this study possibly had the appropriate amount of exercise for enhancing SOD activity. Especially, given that middle-aged and aged horses at Japan Horseback Riding Club had more chances of riding lessons because of their manageability, their activity level could always be sufficient to retain SOD, as opposed to the young horses. Furthermore, the involvement of ADN in oxidative stress should be noted as well. Oxidative stress may be downregulated by ADN, resulting in an increase of SOD and a decrease of MDA (Wang et al. 2014). Although the definite correlation between ADN and oxidative stress was not confirmed in this study, different analysis such as molecular biological approaches should be applied to dig deeper into this matter. This study has several limitations. First, a small number of healthy horses were available at Japan Horseback Riding Club, so we had a limited number of animals in our study. Second, since the animals involved in the study were randomly selected, there was no gender matching performed. Third, for the same reason as second, the classification of the age groups may not be appropriate. Fourth, all blood samples were collected postprandially, hence postprandial influence cannot be eliminated.

3-5 Conclusion

In conclusion, this study demonstrated that aging might not induce remarkable changes in a metabolic system for horses, possibly due to their continuous daily exercises. As all the horses had constant physical activity on a daily basis, the recovery of ADN level was possibly a confirmation that aged horses could improve their metabolic and immune systems, preventing a negative aging effect. Additionally, they sustained balanced M/L ratio among all the age groups. Moreover, as elevated SOD activities in middle-aged and aged horses were displayed, their exercise protocol was a beneficial strategy to enhance anti-obesity and anti-aging promoters. In fact, due to their continuous daily exercises, the riding horses maintained their physical condition as they grow older.

Aging is an inevitable event for all living organisms, and destruction of metabolic or immune systems accompanied by aging can completely change our lives. Although further research is required for the development of blood indicators, blood analysis can serve as a useful index for early detection and protection from metabolic-related and/or age-related diseases. Finding a key to enhance the metabolic system in order to reduce risks for various age-related diseases can offer new intriguing avenues for a desirable healthy life.

Chapter 4

Future Directions: The Aging Effect in Dolphins

4-1 Introduction

There are many different animal species, studying their biology has often benefited human research and understanding for a long time. Comparative studies, for example, can give us valuable clues for life science and medical fields, and it is useful to establish standard data and discover advantageous features for both animals and humans. We have learned the similarities and differences of the animal metabolic system from this study thus far, so from here, I will describe what we can speculate and what we need for further research.

Given that this comparative study gave us helpful suggestions for metabolism patterns in large animals, it is advantageous to understand the unique metabolism of dolphins. Dolphins are popular aquatic mammals seen in many aquariums around the world, and they are very friendly and active. With their high intelligence, there are various dolphin therapy programs for people as well. In addition to being unique aquatic mammals, the metabolism of dolphins has regularly been studied, and is still an attractive research area today. As I describe below, investigating their hematological relevance to type 2 diabetes in humans may be beneficial for new therapeutic applications. Moreover, dolphins may have an advanced immune system, thus immunological-metabolic crosstalk in dolphins can create an exciting new research aspect.

4-2 Comparative Studies between Dolphins, Horses, and Cows

We spotlighted the lipid and energy metabolism of bottlenose dolphins, Thoroughbred riding horses, and Holstein cows in this study. These 3 different animal species are similarly large animals, and have also been trained or domesticated by humans as well. Regarding physical activity, since horses are quite active, whereas cows exhibit little physical activity, these two animal groups can serve as practical comparisons of dolphins, which also have good motility. In addition, dolphins may be closely related to even-toed ungulates, such as cows, pigs, and camels (Shimamura et al. 1997), so it is intriguing to investigate any similarities and differences between these animal species. Moreover, our laboratory has learned about energy metabolism in horses and cows (Arai et al. 2003; Kimura et al. 2005; Li et al. 2012), thus we can take advantage of this knowledge for this comparative study.

Comparative studies can be helpful because this knowledge can provide essential information, leading to establishing the basal data and discovering the distinct features of the animals. For instance, research on rare species, such as aquatic mammals, is often problematic, thus information about the animals is scant, and the standard values of various blood biomarkers are not established in many cases. Since many different animals are bred in captivity, comparing this knowledge to common and familiar animals can be a valuable strategy in order to investigate and collaborate the information with other rare animal species. Moreover, it is profitable to conduct such comparative studies, contrasting with humans. Advantageous features in animals can often be beneficial for the human medical

field, in order to seek other safe and pragmatic therapeutic options.

4-3 Similarities and Differences between Dolphins, Horses, and Cows

As described previously, there were some similarities between dolphins and horses. First, although it was demonstrated that energy metabolism in each animal group is different, dolphins and horses are both active animals and have good potential for energy production/usage efficiency (Chapter 2). Second, dolphins and horses showed similar LDH isoenzyme patterns, with LDH-3 isoenzyme predominating (Chapter 2). Third, as cholesterol lipoprotein patterns between these 3 animal species suggested, dolphins and horses are HDL-dominant groups and have a distinct fraction of LDL (Chapter 1).

4-4 Uniqueness of the Metabolic System in Dolphins

Dolphins are well known for being highly intelligent marine mammals. Since they have a large brain like humans, it is hypothesized that dolphins have high blood glucose carrying capacity in their blood to support their large brain (Goodwin 1956). However, dolphins mainly eat fish, which is a high-protein diet, so they needed to evolve their metabolic system to obtain enough sugar from their diet. For humans, a high-protein diet is effective for the treatment of type 2 diabetes (Gannon et al. 2003), but it also has a negative impact on renal health. It was reported that Atlantic bottlenose dolphins (*Tursiops truncatus*) have a prolonged glucose tolerance curve and naturally maintain a fasting hyperglycemia, which is found in diabetes mellitus in humans (Ridgeway 1972; Ridgeway et al. 1970). Interestingly,

it appeared that dolphins have advanced insulin resistance without harming renal function (Venn-Watson et al. 2011).

Since dolphins have a unique metabolism, some researchers insisted the possibility that dolphins may be an ideal animal model for type 2 diabetes. Although there are similarities between dolphins and humans, dolphins may have a very different metabolic system, such as energy metabolism. Given that dolphins are aquatic mammals and have a layer of blubber, it is possible to have different energy requirements and a different storing system. However, it is beneficial to understand their metabolic system, and investigating how dolphins metabolize a high-protein diet and maintain their health can guide us to a new avenue for novel therapeutic options.

4-5 Further Research: The Aging Effect in Dolphins

Because of improved health management and veterinary science, older animal populations have been largely expanded these days. However, negative aging effects can impact the metabolic system, and age-related disorders have become one of the major health problems for animals. As it is well known today, since a metabolic system is subject to change with aging, the number of metabolic disorders in animals has been rapidly increasing as well. Prevalence of overweight and obesity is not only for human health problems, but also for animals (Tvarijonaviciute et al. 2012; Martin et al. 2014; Robin et al. 2015). These disorders are a critical issue for both animals and humans, and its prevention strategies and treatment have been greatly debated all over the world. Therefore, maintaining a

desirable metabolic system can help defeat the negative effects of aging, and is capable of preserving good health throughout life.

As presented in Chapter 3, daily exercises may improve the metabolic system and slow the negative aging effects in riding horses. It appeared that dolphins shared some similarities with horses in terms of energy metabolism patterns, so their daily exercises in the daytime may help maintain their metabolism with increasing age as well. In fact, regarding physical activity, both dolphins and horses have similar patterns, which is managed by humans. The amount of physical activity is maintained constant, and the exercise protocol is always carefully monitored so animals are not stressed. Additionally, in most cases, older dolphins and older horses bred in captivity are clinically healthy, and they can often live longer than the wild condition. Since older animals are often more valuable than younger ones because of their manageability, a more favorable circumstance for exercises can be built with increasing age.

It was reported that hematological and serum chemistry values in aged dolphins are very similar to those in older humans (Venn-Watson et al. 2011). According to this report, aged dolphins demonstrated a normative increasing state of chronic inflammation with age, including hyperglobulinemia, leukocytosis, and neutrophilia. It is well known that the immune system in mammals continually loses its ability to respond properly (Desai et al. 2010). Interestingly, it is believed that marine mammals may have developed a uniquely robust immune system since they have impressive abilities to rapidly heal large wounds, including shark bites, in a microbial challenging environment without any medical attention

(Corkeron et al. 1987). Therefore, exploring an aging dolphin immune system can benefit the health of both dolphins and humans. Furthermore, since it was demonstrated that the dysfunction of a metabolic system is attributed to inflammation (Kanneganti and Dixit 2012), understanding the immunological process that regulates obesity may provide new aspects on the management of various chronic diseases.

4-6 Conclusion

This comparative study suggested that each animal species has a distinct metabolism pattern, but there are some similarities with other species as well. From these similarities, it is possible to presume certain effects on the metabolic system in similar animal groups. Investigating and comparing the metabolism in animals can lead to a better understanding of their normal health condition. Since some animal species are often limited to research, it is important to gain helpful insights from the similarities and the differences with other species.

Evidence now indicates that daily physical activity may help improve the metabolic ability and can attenuate the negative aging effects in active animals, such as riding horses. Although dolphins appear to have a unique metabolic system, it is speculated that their regular physical activity, including shows in the daytime, have a similar impact on their health management when they age. Moreover, dolphins and humans share several features, hence examining the unique metabolic system in dolphins can lead to striking and beneficial theories for new aspects on health and diseases in humans as well.

As stated above, comparative studies can allow us to recognize the remarkable features of the animals. The basic information gained from comparative studies can bring us profitable knowledge for establishing standards of animal basal metabolism. A better understanding of metabolism patterns can lead to more efficient management and disease prevention strategies for these animals.

An abundance of information is hidden in the biology of animals, which we can carefully and ethically exploit for medicine and technology areas. Given that having a balanced metabolic system is necessary for good health, identification of the advantageous metabolic process can bridge a knowledge gap in metabolic-related issues in animals and humans. Generally, the impact of exercise proved to work positively for anti-obesity and anti-aging in animals and humans. In fact, each animal species has a different energy utilization process, thus it is favorable to apply the most appropriate exercise strategy for each animal species. For better health management of animals, it is necessary to understand each metabolic system in order to prevent any dysregulation of metabolism, if it occurs. A comparative study can be the first step to more efficient management and disease prevention strategies. It can also offer new pragmatic avenues for therapeutic approaches for animals and humans.

Summary

In recent years, the progress of veterinary science has contributed to the longevity of animals. However, along this development, the number of metabolic and age-related disorders in animals has been rapidly increasing. Similarly to humans, many major diseases are related to metabolic conditions, therefore investigating the metabolic biomarkers is beneficial for both animals and humans. In this study, we investigated and compared the metabolic systems in large animals, such as dolphins, horses, and cows. Comparative studies are helpful because this knowledge can offer critical information, leading to establishing the basic standard data and discovering the distinct features of the animals. It is also advantageous to use this knowledge for other fields, including the human medical field.

1. Cholesterol Lipoprotein Profiles of Large Animals

Cholesterol profile patterns between dolphins, horses, and cows showed that all animal groups were classified into HDL dominant mammals, such as dogs and cats. Dolphins and horses also displayed clear LDL-Cho peak, which was not shown in cows, thus the cholesterol profile of dolphins is a closer resemblance to that of horses, rather than that of cows. Evidence now indicates that understanding the characteristics in the lipid and lipoprotein parameters of each animal group is necessary for maintaining a healthy metabolism in animals. In addition, various factors can often alter the patterns, so investigating how they are involved in lipid metabolism can lead to a more specific evaluation of an animal's metabolic state.

Overall, monitoring the plasma lipid concentrations and cholesterol profile in animals is useful to detect abnormal metabolic states in order to prevent various metabolic-related disorders.

2. Comparison of Energy Metabolism and LDH Isoenzyme Patterns in Large Animals

Regarding energy metabolism between these animal groups, dolphins appear to have the greatest level of oxidative energy metabolism amongst horses and cows, due to having the greatest levels of plasma MDH activity. In addition, dolphin energy production/usage efficiency was second (M/L ratio = 0.67) behind that of horses (M/L ratio = 0.79). Overall, these results suggest that dolphins may possibly generate more energy than horses, especially for increased mobility in the water; however, all of the produced energy is not utilized for various reasons, such as a smaller tank size or other environmental limitations. However, cows demonstrated the highest plasma LDH activity amongst all animal species. This may have been attributed to their lactating state. Although all animal groups displayed a different plasma LDH isoenzyme pattern distribution, dolphins and horses demonstrated a similarity with LDH-3 isoenzyme predominating in plasma, as opposed to LDH-1 in cows, which would reflect “Symmorphosis” of these two species and their aerobic/anaerobic energy metabolism needs.

Overall, plasma MDH and LDH activity levels, M/L ratio, and plasma LDH isoenzyme pattern can all be useful indicators for a better understanding of the oxidative energy metabolism and monitoring of a captive animals' health. As it is

not easy to obtain tissue samples from animals, the development of blood indicators for evaluating the whole body metabolic state is necessary, and further research is required and should be pursued.

3. The Aging Effect on the Metabolic System in Riding Horses

The study about the aging effect demonstrated that aging might not induce remarkable changes in a metabolic system for horses, possibly due to their continuous daily exercises. As all the horses had constant physical activity on a daily basis, recovery of the ADN level was possibly a confirmation that aged horses could improve their metabolic and immune systems, preventing a negative aging effect. Additionally, they sustained a balanced M/L ratio among all the age groups. Moreover, as elevated SOD activities in middle-aged and aged horses were displayed, their exercise protocol was a beneficial strategy to enhance anti-obesity and anti-aging promoters. In fact, due to their continuous daily exercises, the riding horses maintained their physical condition as they grew older.

Aging is an inevitable event for all living organisms. The destruction of the metabolic or immune systems accompanied with aging can completely change our lives. Although further research is required for the development of blood indicators, blood analysis can serve as a useful index for early detection and protection from metabolic-related and/or age-related diseases. Finding a key to enhance the metabolic system in order to reduce the risks for various age-related diseases can offer new intriguing avenues for a desirable and healthy life.

4. Future Directions: The Aging Effect in Dolphins

This comparative study suggested that each animal species has a distinct metabolism pattern, but there are some similarities with other species as well. From these similarities, it is possible to presume certain effects on the metabolic system in similar animal groups. Investigating and comparing the metabolism in animals can lead to a better understanding of their normal health condition. Since some animal species are often limited to research, it is important to gain helpful insights from the similarities and the differences with other species.

Evidence now indicates that daily physical activity may help improve the metabolic ability and can attenuate the negative aging effects in active animals, such as riding horses. Although dolphins appear to have a unique metabolic system, it is speculated that their regular physical activity, including shows in the daytime, have a similar impact on their health management when they age. Moreover, dolphins and humans share several features, hence examining the unique metabolic system in dolphins can lead to new, impactful, and beneficial health and disease theories for humans as well.

As stated above, comparative studies can allow us to recognize the remarkable features of the animals. The basic information gained from comparative studies can bring us profitable knowledge for establishing the standards of animal basal metabolism. A better understanding of metabolism patterns can lead to more efficient management and disease prevention strategies for these animals.

An abundance of information is hidden in the biology of animals, which we can

carefully and ethically exploit for medicine and technology areas. Given that having a balanced metabolic system is necessary for good health, identification of the advantageous metabolic process can bridge a knowledge gap in metabolic-related issues in animals and humans. Generally, the impact of exercise proved to work positively for anti-obesity and anti-aging in animals and humans. In fact, each animal species has a different energy utilization process, thus it is favorable to apply the most appropriate exercise strategy for each animal species. To improve the health management of animals, it is necessary to understand each metabolic system in order to prevent any dysregulation of metabolism, if it occurs. A comparative study can be the first step to more efficient management and disease prevention strategies. It can also offer new pragmatic avenues and therapeutic approaches for animals and humans.

References

- Arai T, Inoue A, Uematsu Y, Sako T, Kimura N. Activities of enzymes in the malate-aspartate shuttle and the isoenzyme pattern of lactate dehydrogenase in plasma and peripheral leukocytes of lactating Holstein cows and riding horses. *Res Vet Sci* (2003) 75 (1): 15-19
- Arai T, Machida T, Sasaki M, Oki Y. Hepatic enzyme activities and plasma insulin concentrations in diabetic herbivorous voles. *Vet Res Commun* (1998) 13 (6): 421-426
- Arai T, Takahashi M, Araki K, Washizu T. Activities of enzymes related to the malate-aspartate shuttle in the blood cells of thoroughbreds undergoing training exercise. *Vet Res Commun* (2001) 25 (7): 577-583
- Asadi F, Mohri M, Adibmoradi M, Pourkabir M. Serum lipid and lipoprotein parameters of Turkman horses. *Vet Clin Pathol* (2006) 35 (3): 332-334
- Asper ED, Cornell LH, Duffield DA, *et al.* Hematology and serum chemistry values in bottlenose dolphins. In: Leatherwood S, Reeves RR, Eds *The Bottlenose Dolphin*. New York, NY: Academic Press (1990) pp. 479-485
- Bell AW, Bauman DE. Adaptations of glucose metabolism during pregnancy and lactation. *J Mammary Gland Biol Neoplasia* (1997) 2 (3): 265-278
- Bergmeyer HU, Brent E. Malate dehydrogenase UV assay. In: Bergmeyer HU, Ed. *Methods of enzymatic analysis, Vol.2*. Verlag Chemie Weinheim, ed. New York, NY: Academic Press (1974) pp. 613-617
- Bossart GD, Reidarson TH, Dierauf LA, Duffield DA. Clinical pathology. In: Gulland FMD, Dierauf LA. Eds. *CRC Handbook of Marine Mammal Medicine*, 2nd ed. Washington, DC: CRC Press (2001) pp. 383-436
- Boyd J, West JW, Bernard JK. Effects of the addition of direct-fed microbials and glycerol to the diet of lactating dairy cows on milk yield and apparent efficiency of yield. *J Dairy Sci* (2011) 94 (9): 4616-4622
- Bray GA, Bellanger T. Epidemiology, trends, and morbidities of obesity and the metabolic syndrome. *Endocrine* (2006) 29 (1): 109-117

Brochu-Gaudreau K, Rehfeldt C, Blouin R, Bordignon V, Murphy BD, Palin MF. Adiponectin action from head to toe. *Endocrine* (2010) 37 (1): 11-32

Carroll CL, Huntington PJ. Body condition scoring and weight estimation of horses. *Equine Vet J* (1988) 20 (1): 41-45

Chapman MJ. Comparative analysis of mammalian plasma lipoproteins. In: Segrest JP and Alberts JJ (eds) *Plasma Lipoprotein*. Academic Press, Boston, *Methods in Enzymology* (1986) Vol. 128A, 70–143

Corkeron PJ, Morris RJ, Bryden MM. A note on healing of large wounds in bottlenose dolphins, *Tursiops truncatus*. *Aquat Mamm* (1987) 13.3: 96-98

Craik JD, Young JD, Cheeseman CI. GLUT-1 mediation of rapid glucose transport in dolphin (*Tursiops truncatus*) red blood cells. *Am J Physiol* (1998) 274 (1 Pt 2): R112-R119

Dawson DM, Goodfriend TK, Kaplan NO. Lactic dehydrogenase: Functions of the two types. *Science* (1964) 143 (3609): 929-933

de Heredia FP, Gómez-Martínez S, Marcos A. Obesity, inflammation and the immune system. *Proc Nutr Soc* (2012) 71 (2): 332-338

Desai A, Grolleau-Julius A, Yung R. Leukocyte function in the aging immune system. *J Leukoc Biol* (2010) 87 (6): 1001-1009

Duren SE. Feeding the endurance horse. In: *Advances in Equine Nutrition*, Ed. Pagan, JD, Nottingham, UK: Nottingham University Press (1998) pp. 351-363

Duvivier BMFM, Schaper NC, Bremers MA, van Crombrugge G, Menheere PPCA, Kars M, Savelberg HHCM. Minimal intensity physical activity (standing and walking) of longer duration improves insulin action and plasma lipids more than shorter periods of moderate to vigorous exercise (cycling) in sedentary subjects when energy expenditure is comparable. *PloS One* (2013) 8 (2): e55542

Fair PA, Hulsey TC, Varela RA, *et al.* Hematology, serum chemistry, and cytology findings from apparently healthy Atlantic Bottlenose Dolphins (*Tursiops truncatus*) inhabiting the estuarine waters of Charleston, South Carolina. *Aquat Mamm* (2006) 32 (2): 182-195

Ford RB. Clinical management of lipemic patients. *Compend Contin Educ Dent* (1996) 18 (10): 1053–1060

Frank N, Andrews FM, Elliott SB, Lew J, Boston RC. Effects of rice bran oil on plasma lipid concentrations, lipoprotein composition, and glucose dynamics in mares. *J Anim Sci* (2005) 83 (11): 2509-2518

Gannon MC, Nuttall FQ, Saeed A, Jordan K, Hoover H. An increase in dietary protein improves the blood glucose response in persons with type 2 diabetes. *Am J Clin Nutr* (2003) 78 (4): 734-741

Garland Jr T, Huey R. Testing symmorphosis: Does structure match functional requirements? *Evolution* (1987) 41 (6): 1404-1409

Gawel S, Wardas M, Niedworok E, Wardas P. Malondialdehyde (MDA) as a lipid peroxidation marker. *Wiad Lek* (2004) 57 (9-10): 453-455

Goodwin RF. The distribution of sugar between red cells and plasma: variations associated with age and species. *J Physiol* (1956) 134 (1): 88-101

Gregor MF, Hotamisligil GS. Inflammatory mechanisms in obesity. *Annu Rev Immunol* (2011) 29: 415-445

Harman D. Aging: a theory based on free radical and radiation chemistry. *J Gerontol* (1956) 11 (3): 298-300

Hedekov J, Capito K, Thams P. Cytosolic ratios of free [NADPH]/[NADP⁺] and [NADH]/[NAD⁺] in mouse pancreatic islets, and nutrient-induced insulin secretion. *Biochem J* (1987) 241 (1): 161-167

Hirakawa Y, Kawasumi K, Lee P, Mori N, Yamamoto I, Terasawa F, Arai T. Determination of oxidative energy metabolism and plasma LDH isoenzyme patterns of dolphins. *Open Vet Sci J* (2012) 6: 30-36

Jericó MM, De Camargo CF, Kajihara K, Moreira MA, Gonzales R, Machado FL, Nunes VS, Catanozi S, Nakandakare ER. Chromatographic analysis of lipid fractions in healthy dogs and dogs with obesity or hyperadrenocorticism. *J Vet Diagn Invest* (2009) 21 (2): 203-207

Johnson MC. Hyperlipidemia disorders in dogs. *Compend Contin Educ Dent* (2005) 27 (5) 361-364

Jorgensen SB, Rose AJ. How is AMPK activity regulated in skeletal muscles during exercise? *Front Biosci* (2008) 13: 5589-604

Kaloustian HD, Stolezenbach FE, Everse J, Kaplan NO. Lactate dehydrogenase of lobster (*Hornarus americanus*) tail muscle I. Physical and chemical properties. J Biol Chem (1969) 244 (11): 2891-901

Kanneganti TD, Dixit VD. Immunological complications of obesity. Nat Immunol (2012) 13 (8): 707-712

Kasamatsu M, Kawauchi R, Tsunokawa M, *et al.* Comparison of serum lipid compositions, lipid peroxide, α -tocopherol and lipoproteins in captive marine mammals (bottlenose dolphins, spotted seals and West Indian manatees) and terrestrial mammals. Res Vet Sci (2009) 86 (2): 216-222

Kasamatsu M, Tsunokawa M, Taki M, Higuchi H, Nagahata H. Serum lipid peroxide and α -tocopherol concentrations and superoxide dismutase activity in captive bottle-nosed dolphins. Am J Vet Res (2001) 62 (12): 1952-1956

Kawasumi K, Yamamoto M, Koide M, Okada Y, Mori N, Yamamoto I, Arai T. Aging effect on plasma metabolites and hormones concentrations in riding horses. Open Vet J (2015) 5 (2): 154-157

Kimura N, Yoshimura I, Sako T, Inoue A, Tadami K, Arai T. Changes in activities of enzymes related to energy metabolism in peripheral leukocytes of fattening steers. Vet Res Commun (2005) 29 (1): 19-26

Laflamme DP. Companion Animals Symposium: Obesity in dogs and cats: What is wrong with being fat? J Anim Sci (2012) 90 (5): 1653-1662

Watson TDG, Barrie J. Lipoprotein metabolism and hyperlipidaemia in the dog and cat: A review. J Small Anim Pract (1993) 34 (10): 479-487

Latner AL, Siddiqui SA, Skillen AW. Pyruvate inhibition of lactate dehydrogenase activity in human tissue extracts. Science (1966) 154 (3748): 527-529

Lauris V, Cahill GF. Hepatic glucose phosphotransferases. Variations among species. Diabetes (1966) 15 (7): 475-479

Li G, Lee P, Mori N, Yamamoto I, Arai T. Long term intensive exercise training leads to a higher plasma malate/lactate dehydrogenase (M/L) ratio and increased level of lipid mobilization in horses. Vet Res Commun (2012) 36 (2): 149-155

Li XD, Sun GF, Zhu WB, Wang YH. Effects of high intensity exhaustive exercise on SOD, MDA, and NO levels in rats with knee osteoarthritis. *Genet Mol Res* (2015) 14 (4): 12367-12376

Lira FS, Pimentel GD, Santos RV, Oyama LM, Damaso AR, Oller do Nascimento CM, Viana VA, Boscolo RA, Grassmann V, Santana MG, Esteves AM, Tufik S, de Mello MT. Exercise training improves sleep pattern and metabolic profile in elderly people in a time-dependent manner. *Lipids Health Dis* (2011) 10 (113): 1-6

Lu JY, Huang KC, Chang LC, Huang YS, Chi YC, Su TC, Chen CL, Yang WS. Adiponectin: a biomarker of obesity-induced insulin resistance in adipose tissue and beyond. *J Biomed Sci* (2008) 15 (5): 565–576

Lukens JR, Dixit VD, Kanneganti TD. Inflammasome activation in obesity-related inflammatory diseases and autoimmunity. *Discov Med* (2011) 12 (62): 65-74

Marino L. Quantifying brain-behavior relations in cetaceans and primates. *Trends Ecol Evol* (1998) 13 (10): 408

Martin LJ, Lutz TA, Daumas C, Bleis P, Nguyen P, Biourge V, Dumon HJ. Acute hormonal response to glucose, lipids and arginine infusion in overweight cats. *J Nutr Sci* (2014) 3: e8

Mori N, Lee P, Kondo K, Kido T, Saito T, Arai T. Potential use of cholesterol lipoprotein profile to confirm obesity status in dogs. *Vet Res Commun* (2011) 35 (4): 223-235

Muranaka S, Mori N, Hatano Y, Saito TR, Lee P, Kojima M, Kigure M, Yagishita M, Arai T. Obesity induced changes to plasma adiponectin concentration and cholesterol lipoprotein composition profile in cats. *Res Vet Sci* (2011) 91 (3): 358-361

Osborn O, Olefsky JM. The cellular and signaling networks linking the immune system and metabolism in disease. *Nat Med* (2012) 18 (3): 363-374

Praticò D. Lipid peroxidation and the aging process. *Sci Aging Knowledge Environ* (2002) 2002 (50): re5

Ridgeway SH. *Mammals of the sea: biology and medicine*. Springfield (IL): Charles C Thomas. (1972) p690-747

Ridgway SH, Simpson JG, Patton GS, Gilmartin. Hematologic findings in certain small cetaceans. *J Am Vet Med Assoc* (1970) 157(5):566-575

Robin CA, Ireland JL, Wylie CE, Collins SN, Verheyen KL, Newton JR. Prevalence of and risk factors for equine obesity in Great Britain based on owner-reported body condition scores. *Equine Vet J* (2015) 47 (2): 196-201

Rommel SA, Lowenstine LJ. Gross and microscopic anatomy. In: Gulland FMD, Dierauf LA, Eds. *CRC Handbook of Marine Mammal Medicine*. 2nd ed. Washington DC: CRC Press (2001) pp. 129-164

Ruderman NB, Carling D, Prentki M, Cacicedo JM. AMPK, insulin resistance, and the metabolic syndrome. *J Clin Invest* (2013) 123 (7): 2764-2772

Sako T, Urabe S, Kusaba A, Kimura N, Yoshimura I, Tazaki H, Imai S, Ono K, Arai T. Comparison of plasma metabolite concentrations and lactate dehydrogenase activity in dogs, cats, horses, cattle and sheep. *Vet Res Commun* (2007) 31 (4): 413-417

Sasaki T, Nakata R, Inoue H, Shimizu M, Inoue J, Sato R. Role of AMPK and PPAR γ 1 in exercise-induced lipoprotein lipase in skeletal muscle. *Am J Physiol Endocrinol Metab* (2014) 306 (9): E1085-1092

Setoyama C, Joh T, Tshuzuki T, Shimada K. Structural organization of the mouse cytosolic malate dehydrogenase gene: comparison with that of the mouse mitochondrial malate dehydrogenase gene. *J Mol Biol* (1988) 202 (3): 355-364

Shimamura M, Yasue H, Ohshima K, *et al.* Molecular evidence from retroposons that whales form a clade within even-toed ungulates. *Nature* (1997) 388 (6643): 666-670

Stienstra R, Tack CJ, Kanneganti TD, Joosten LA, Netea MG. The inflammasome puts obesity in the danger zone. *Cell Metab* (2012) 15 (1): 10-18

Storey KB, Hochachka PW. Glycolytic enzymes in muscle of the Pacific dolphin: role of pyruvate kinase in aerobic-anaerobic transition during diving. *Comp Biochem Physiol B* (1974) 49 (1B): 119-128

Suominen H, Heikkinen E. Enzyme activities in muscle and connective tissue of *M. vastus lateralis* in habitually training and sedentary 33 to 70-year-old men. *Eur J Appl Physiol Occup Physiol* (1975) 34 (4): 249-254

Taylor CR, Weibel ER. Design of the mammalian respiratory system. I. Problem and strategy. *Respir Physiol* (1981) 44 (1): 1-10

Terasawa F, Kitamura M. Hyperlipemia of captive bottlenose dolphins during pregnancy. *J Vet Med Sci* (2005) 67 (3): 341-344□

Terasawa F, Kitamura M, Fujimoto A, Hayama S. Influence of diet on hematological characteristics in bottlenose dolphins. *Jpn J Zoo Wildl Med* (1999) 4 (2): 117-124

Terasawa F, Kitamura M, Fujimoto A, Hayama S. Seasonal changes of blood composition in captive bottlenose dolphins. *J Vet Med Sci* (2002) 64 (11): 1075-1078

Terpstra AH, Sanchez-Muniz FJ, West CE, Woodward CJ. The density profile and cholesterol concentration of serum lipoproteins in domestic and laboratory animals. *Comp Biochem Physiol B* (1982) 71 (4): 669-673

Tvarijonaviciute A, Ceron JJ, Holden SL, Cuthbertson DJ, Biourge V, Morris PJ, German AJ. Obesity-related metabolic dysfunction in dogs: a comparison with human metabolic syndrome. *BMC Vet Res* (2012) 8: 147

Venn-Watson S, Carlin K, Ridgway S. Dolphins as animal models for type 2 diabetes: sustained, post-prandial hyperglycemia and hyperinsulinemia. *Gen Comp Endocrinol* (2011) 170 (1): 193-199

Venn-Watson S, Smith CR, Gomez F, Jensen ED. Physiology of aging among healthy, older bottlenose dolphins (*Tursiops truncatus*): comparisons with aging humans. *J Comp Physiol B* (2011) 181 (5): 667-680

Wang X, Pu H, Ma C, Jiang T, Wei Q, Zhang C, Duan M, Shou X, Su L, Zhang J, Yang Y. Adiponectin Abates Atherosclerosis by Reducing Oxidative Stress. *Med Sci Monit* (2014) 20: 1792-1800

Washizu T, Kuramoto E, Abe M, Sako T, Arai T. A comparison of the activities of certain enzymes related to energy metabolism in leukocytes in dogs and cats. *Vet Res Commun* (1998) 22 (3): 187-192

Washizu T, Takahashi M, Azakami D, Ikeda M, Arai T. Activities of enzymes in the Malate-Aspartate shuttle in the peripheral leukocytes of dogs and cats. *Vet Res Commun* (2001) 25 (8): 623-629

Watson TDG, Barrie J. Lipoprotein metabolism and hyperlipidemia in the dog and cat— a review. *J Small Anim Pract* (1993) 34 (10): 479-487

Watson TD, Burns L, Love S, Packard CJ, Shepherd J. The isolation, characterisation and quantification of the equine plasma lipoproteins. *Equine Vet J* (1991) 23 (5): 353-359

Weibel ER, Taylor RC. Design of the mammalian respiratory system. *Respir Physiol* (1981) 44: 1-164

Whitney MS. Evaluation of hyperlipidemias in dogs and cats. *Semin Vet Med Surg (Small Anim)* (1992) 7 (4): 292–300

Williams TM, Haun JE, Friedl WA, Hall RW, Bivens LW. Assessing the thermal limits of bottlenose dolphins: a cooperative study by trainers, scientists and animals. *IMATA Soundings Fall* (1992) 16-17

Yamauchi T, Kamon J, Minokoshi Y, Ito Y, Waki H, Uchida S, Yamashita S, Noda M, Kita S, Ueki K, Eto K, Akanuma Y, Froguel P, Foufelle F, Ferre P, Carling D, Kimura S, Nagai R, Kahn BB, Kadowaki T. Adiponectin stimulates glucose utilization and fatty-acid oxidation by activating AMP-activated protein kinase. *Nat Med* (2002) 8 (11): 1288-1295

Zhang BB, Zhou G, Li C. AMPK: an emerging drug target for diabetes and the metabolic syndrome. *Cell Metab.* (2009) 9 (5): 407-416

Acknowledgements

The authors involved in the studies above wish to thank Enoshima Aquarium, Koizumi Milk Farm, YCL Horse Clinic and Nippon Veterinary and Life Science University Equestrian Club for their help with collecting blood samples from the animals and their valuable advice. The authors also wish to thank the owners of the chinchillas and the laboratories for providing the specimens. The comparative biochemistry studies on dolphins and horses (Chapter 1-3) were supported in part by the Strategic Research Base Development Program for Private Universities from the Ministry of Education, Culture, Sports, Science and Technology of Japan (MEXT), 2008-2012, and Grant-in-Aid for Scientific Research (No. 21380195) from the MEXT.

I thank all the members of the division of Veterinary Biochemistry in Nippon Veterinary and Life Science University for the discussions and help in preparing the manuscript.